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#### 23 RISPERIDONE QUANTITATION AND CONFIRMATION BY LCMS

# 23.1 Summary

23.1.1 Risperidone and its proprietary internal standard, R68808, are extracted from biological samples with an acetonitrile precipitation and analyzed by high performance liquid chromatography-electrospray ionization mass spectrometry (LC-ESI-MS).

## 23.2 Specimen Requirements

23.2.1 One mL blood, fluid or tissue homogenate.

# 23.3 Reagents and Standards

- 23.3.1 Ammonium acetate
- 23.3.2 Acetic Acid
- 23.3.3 Methanol
- 23.3.4 Acetonitrile
- 23.3.5 Risperidone (Janssen Pharmaceuticals)
- 23.3.6 R68808 (Janssen Pharmaceuticals, internal standard)

## 23.4 Solutions, Internal Standard, Calibrators and Controls

- 23.4.1 5 mM Ammonium Acetate: Weight 0.19 g ammonium acetate. Transfer to 500 mL volumetric flask and QS to volume with dH<sub>2</sub>O
- 23.4.2 Working standard solution for risperidone (0.01 mg/mL)
  - 23.4.2.1 Pipet 100 μl of 1 mg/mL stock solution of risperidone into a 10 mL volumetric flask and QS to volume with acetonitrile
- 23.4.3 Quality Control (QC) standard solution of risperidone (0.01 mg/mL)
  - 23.4.3.1 Pipet 100 µl of separate 1 mg/mL stock solution of risperidone (different preparation than calibrators) into a 10 mL volumetric flask and QS to volume with acetonitrile
- 23.4.4 Internal standard working solution
  - 23.4.4.1 0.01 mg/mL R68808: Pipet 100  $\mu$ L of 1 mg/mL R68808 stock solution into 10 mL volumetric flask and QS to volume with acetonitrile
- 23.4.5 To prepare the calibration curve, pipet the following volumes of the 0.01 mg/mL risperidone working solution into appropriately labeled 16 x 125 mm screw cap test tubes. Add 1 mL blank blood to obtain the final concentrations listed below.

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Amount of Standard ( $\mu L$ )	Final concentration of risperidone (mg/L)
100	1
50	0.5
20	0.2
10	0.1
5	0.05

- 23.4.6 Controls
  - 23.4.6.1 Risperidone Control
    - 23.4.6.1.1 Pipet 50  $\mu$ L of the 0.01 mg/mL risperidone QC solution into an appropriately labeled tube. Add 1 mL blank blood to achieve final concentration of 0.5 mg/L.
  - 23.4.6.2 Negative control. Blood bank blood or equivalent determined not to contain risperidone.

# 23.5 Apparatus

- 23.5.1 Test tubes, 16 x 125 mm, round bottom, borosilicate glass with Teflon caps
- 23.5.2 Test tubes, 16 x 114 mm, glass centrifuge, conical bottom
- 23.5.3 Centrifuge capable of 2000-3000 rpm
- 23.5.4 Nitrogen evaporator with heating block
- 23.5.5 Vortex mixer
- 23.5.6 GC autosampler vials with inserts
- 23.5.7 LC/MS: Agilent Model 1100 LC-MSD
  - 23.5.7.1 LCMS Instrument Conditions. The following instrument conditions may be modified to adjust or improve separation and sensitivity.
    - 23.5.7.1.1 Elution conditions:
      - 23.5.7.1.1.1 Column: Agilent Hypersil BDS 125 mm X 3 mm, 3 µM particle size
      - 23.5.7.1.1.2 Column thermostat: 35° C
      - 23.5.7.1.1.3 Solvent A: 5 mM ammonium acetate (500 mL) containing 150 μL acetic acid
      - 23.5.7.1.1.4 Solvent B: acetonitrile
      - 23.5.7.1.1.5 Isocratic elution, stop time: 4.00 min

Time	Solv. B	Flow
0.00	80	0.6

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23.5.7.1.2 Spray Chamber

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 $\begin{array}{lll} 23.5.7.1.2.1 & Ionization Mode: Electrospray \\ 23.5.7.1.2.2 & Gas Temperature: 300^{\circ} C \\ 23.5.7.1.2.3 & Drying Gas (N_2): 11.8 L/min \\ 23.5.7.1.2.4 & Nebulizer pressure: 30 psig \\ 23.5.7.1.2.5 & Vcap (Positive): 2500 V \\ \end{array}$ 

## 23.5.7.1.3 Selected Ion Monitoring (quantitation ions)

Time	Group Name	SIM	Frag-	Gain	SIM	Actual
(min)		Ion	Mentor	EMV	Resol.	Dwell
0.00	risperidone	191	115	1.0	High	114
		411				114
		<u>412</u>				114
	RS68808	201				114
		421				114

23.5.7.1.3.1 Polarity: Positive 23.5.7.1.3.2 Injection volume: 4 μL

## 23.6 Procedure

- 23.6.1 Label clean 16 x 125 mm screw cap tubes appropriately with calibrators, controls and case sample IDs.
- 23.6.2 Prepare calibrators and controls.
- 23.6.3 Add 1 mL case specimens to the appropriately labeled tubes.
- 23.6.4 Add 50 µL 0.01 mg/mL R68808 internal standard working solution to each tube.
- 23.6.5 Slowly, add dropwise 2 mL cold (freezer temperature) acetonitrile to each tube while vortexing. Continous vortexing, not mere mixing, is essential.
- 23.6.6 Vortex an additional 30 seconds.
- 23.6.7 Place tubes in freezer for at least 30 minutes to facilitate separation.
- 23.6.8 Centrifuge at approximately 2500 rpm for 15 minutes.
- 23.6.9 Transfer top acetonitrile layer to clean conical bottom tubes taking care not to transfer any lower layers.
- 23.6.10 Evaporate to dryness at approximately 50° C under nitrogen.
- 23.6.11 Reconstitute samples in 100 µL acetonitrile. Vortex briefly. Transfer to GC microvials.

### 23.7 Calculation

23.7.1 Drug concentrations are calculated by linear regression analysis using the ChemStation software.

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23.8	Quality Control and Reporting			
	23.8.1 See Toxicology Quality Guidelines			
23.9	REFERENCES			
	23.9.1 J Pearson and R Steiner, in-house development.			